



THE PATCH-CLAMP TECHNIQUE

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WHAT IS A PATCH- CLAMP ?

Applications:

To investigate wide range of electrophysiological cell properties on single cell

Such us:

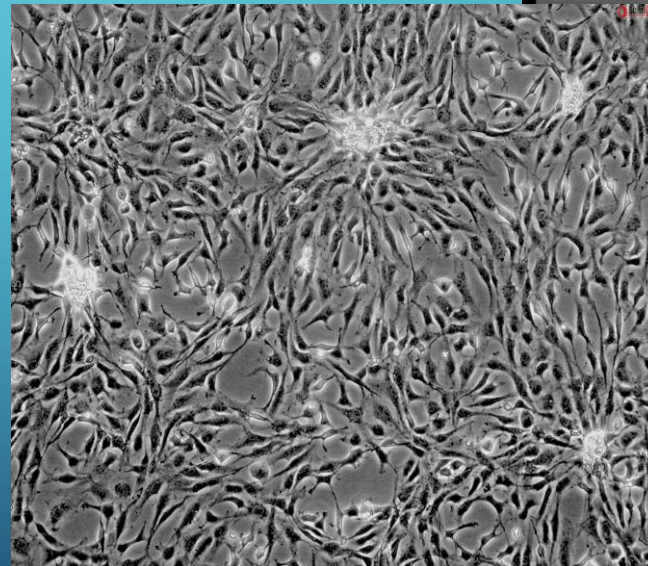
- Current clamp, cell membrane activity and properties (A_p , C_m , Resting membrane potential...)
- Voltage clamp studies on ionic channels (activation, inactivation, voltage dependence, pharmacological blocking....)



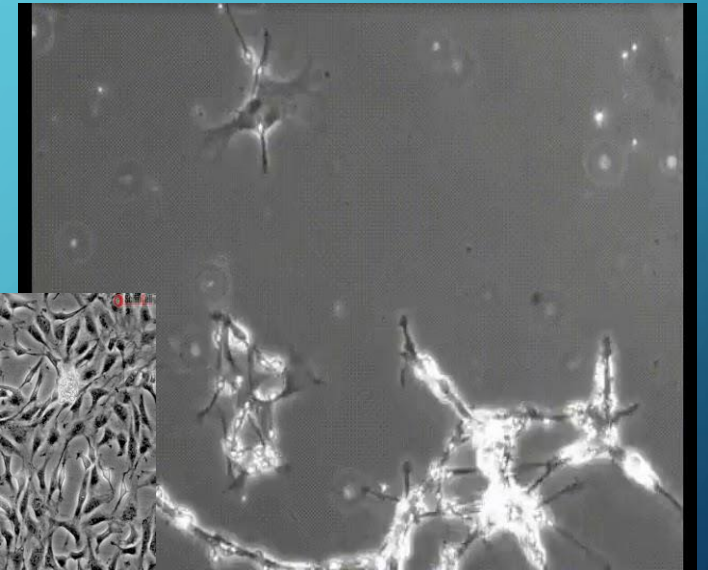
WHAT IS A PATCH- CLAMP ?

Applications:

- Neurons
- Cardiomyocytes
- Muscle fibres
- Pancreatic beta cells
- Kindey cells



Human Cardiac Myocytes



Human Neuron cells

SET-UP

1. Amplifier
2. Data Acquisition Software
3. Microscope
4. Table
5. Faraday cage
6. Imaging/Recording chamber
7. Temperature control
8. Perfusion and Fluid control
9. Stage
10. Micromanipulator
11. Stimulus generator
12. Puller
13. Microforge-grinding
14. Capillary

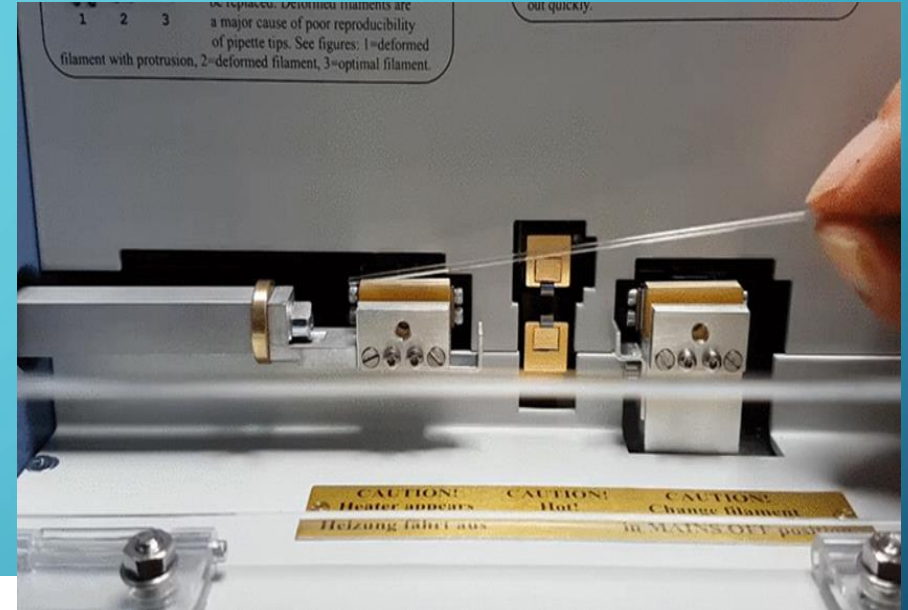


The Puller – How to make your perfect tip ?

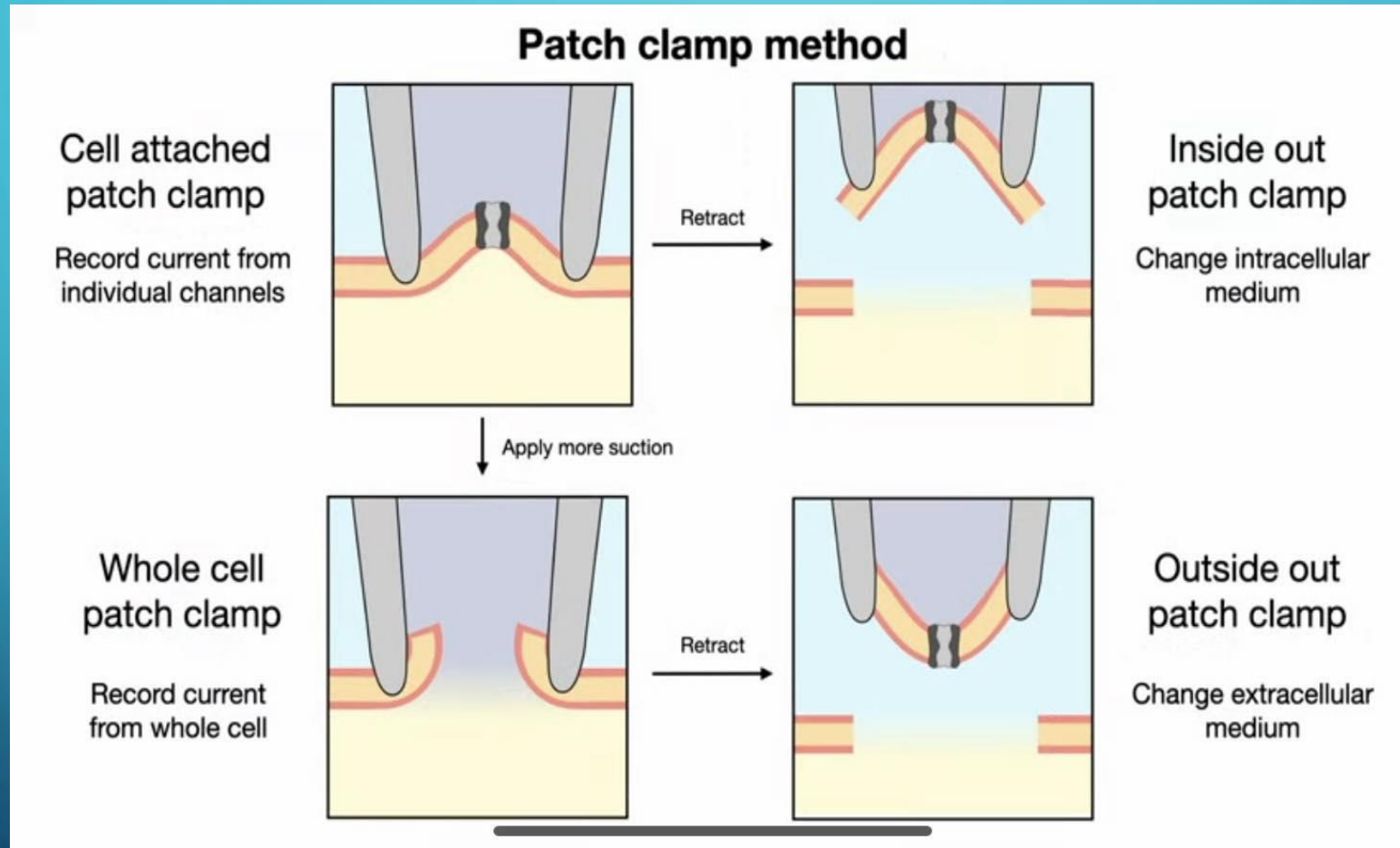
Allows to create the right pipette which is essential for successful recordings and reliable data

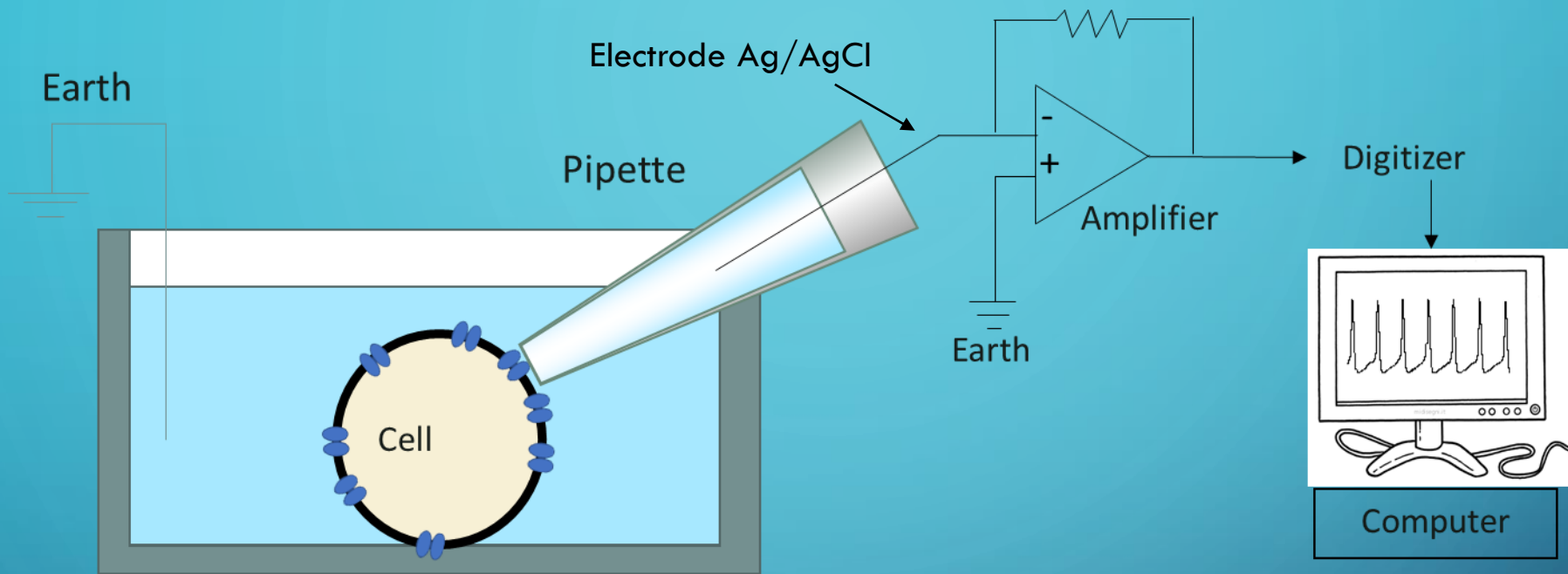
Parameters:

- Temperature
- Velocity
- Applied force
- Filament size

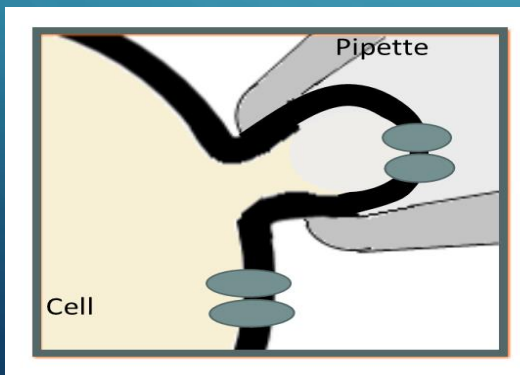


PATCH-CLAMP CONFIGURATIONS





Giga seal

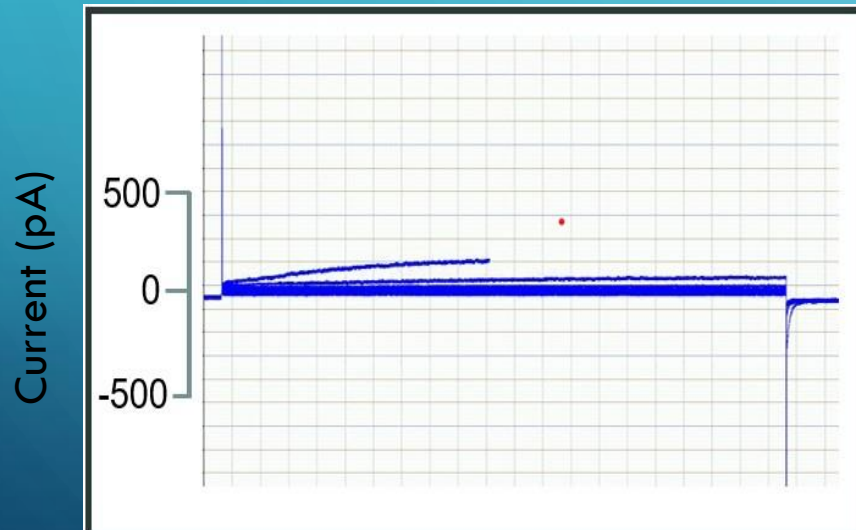


Cell-attached

WHAT CAN WE REGISTER ?

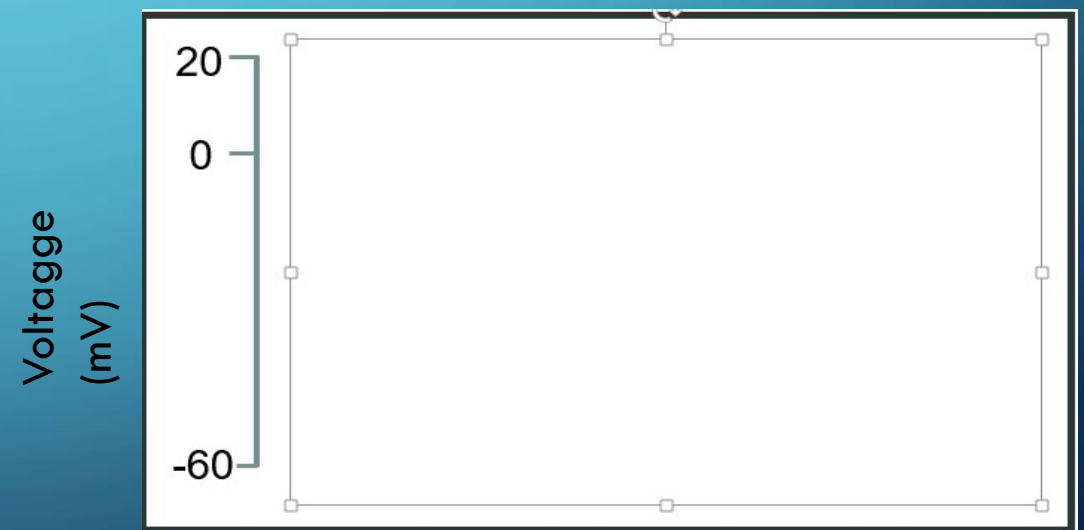
VOLTAGE CLAMP

- impose a membrane voltage
- register the evoked current



CURRENT CLAMP

- impose a current
- register the membrane potential



VOLTAGE CLAMP - WHAT CURRENT ARE WE RECORDING?

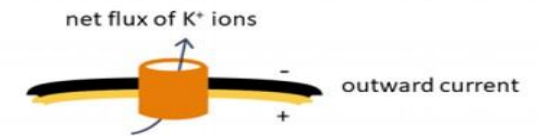
- Direction (inward-outward)
- Kinetics
- Voltage dependence

K⁺ - current

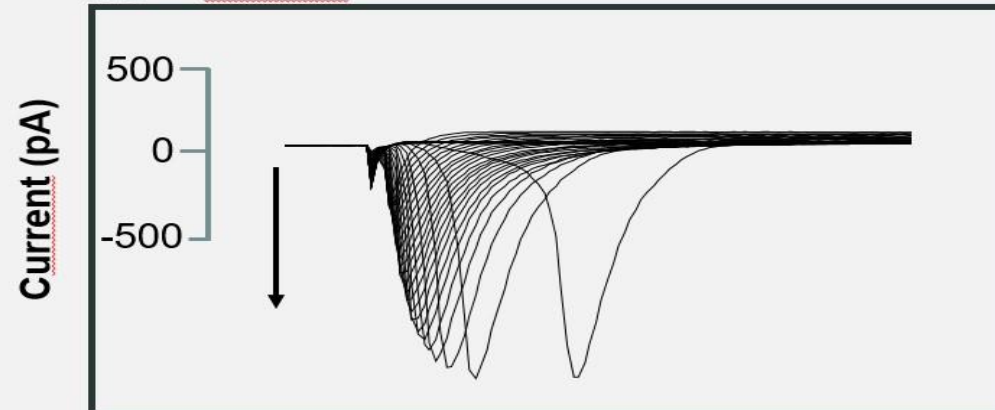


Outward current:

- **Outward positive ions**
- **Inward negative ions**



Na⁺ - current



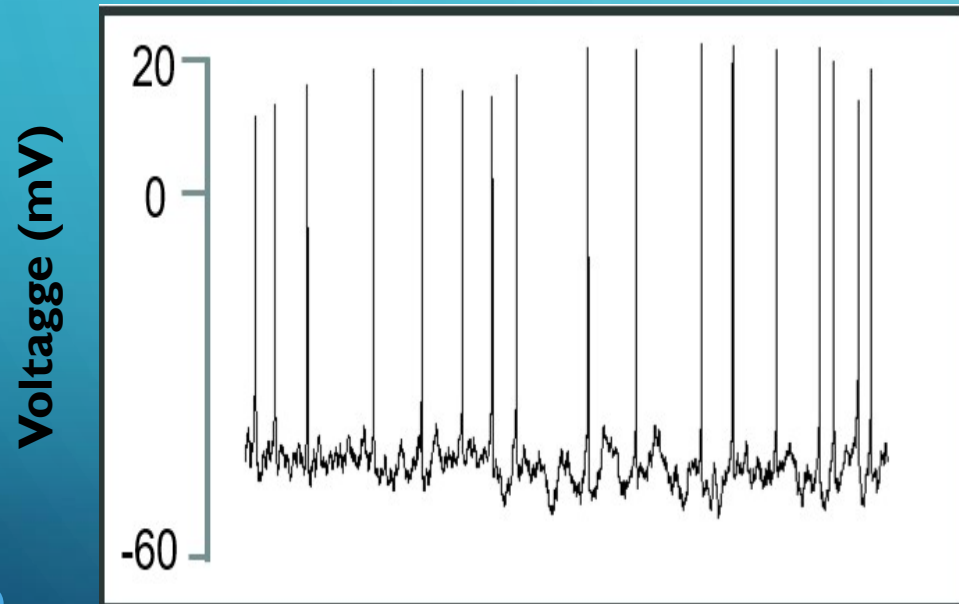
Inward current:

- **Input positive ions**
- **Negative ions in output**

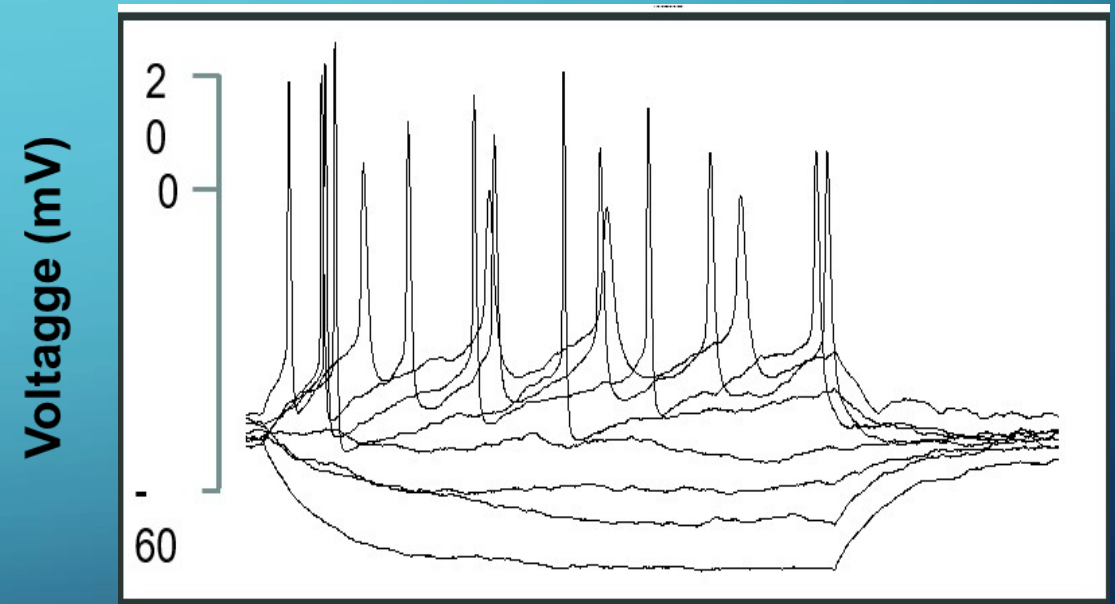


CURRENT CLAMP

- Impose a current
- Register the membrane potential



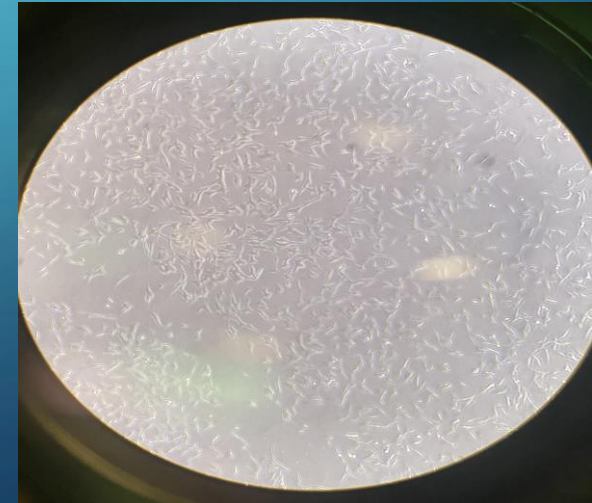
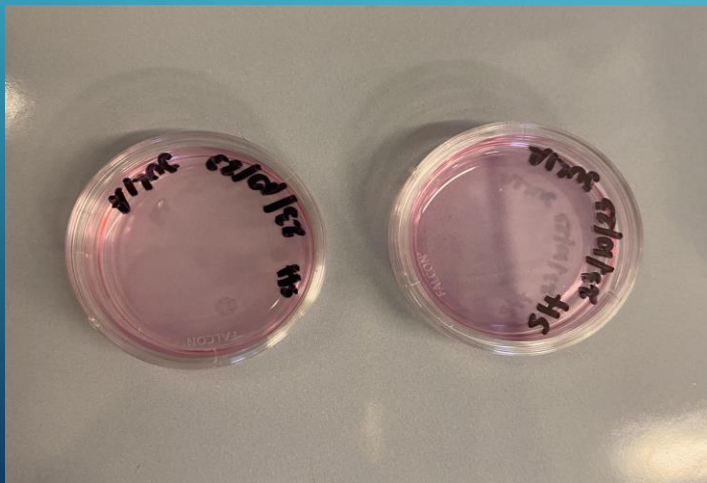
Spontaneous action potentials



Evoked action potentials

PROCEDURE

- Collection of the neuron cells from the T75 flask
- Preparation of the two dishes with 35 mm diameter
- Treatment of the dishes with the Cultrex, incubation for 1 hour
- Removal of Cultrex
- Wash with the medium
- Seeding 125000 cells in each dish with the final volume of the 2ml and adding the medium which contains all the essential components (FBS, Glut, STREEP)
- Conducting the treatment for 7 days
- Obtaining final result of differentiated neurons

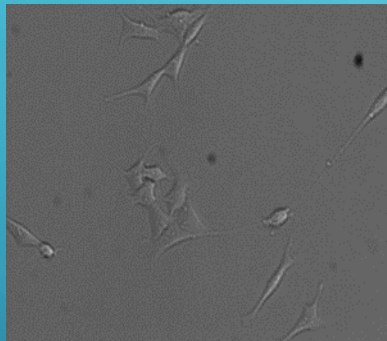


SH cells 3rd day of treatment



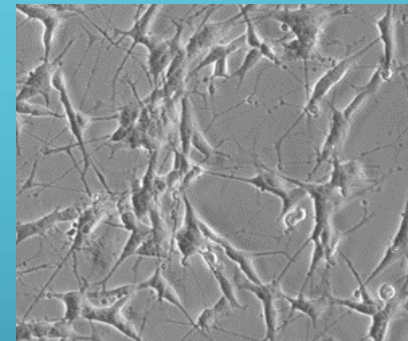
THE CONDUCTED EXPERIMENT

- ❖ TYPE OF CELLS: Undifferentiated SH-SY5Y neuron cells
- ❖ CHARACTERISTICS: SH-SY5Y cells revealed small or even no inward current, but the undifferentiated cells had the highest amplitude of outward currents



Day 1

Undifferentiated neurons



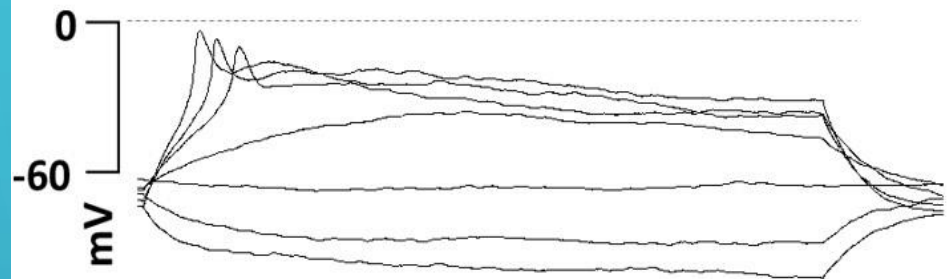
Day 7

Differentiated cells

Medium contained 1 μ M Retinoic Acid - 1%
FBS

SH-SY5Y undifferentiated cells

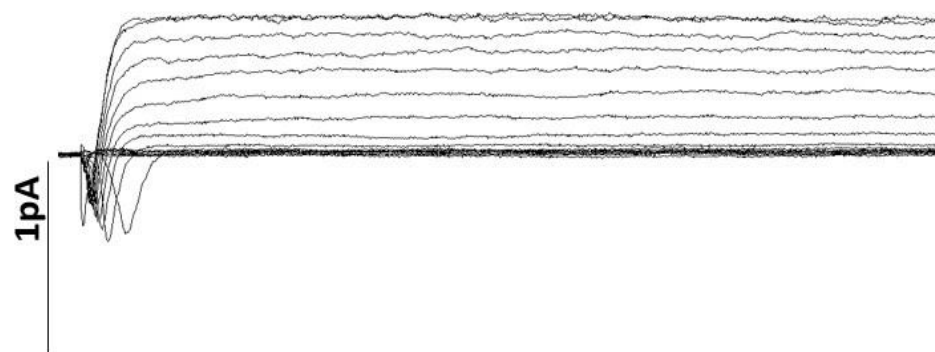
Evoked Action Potential



Resting Potential

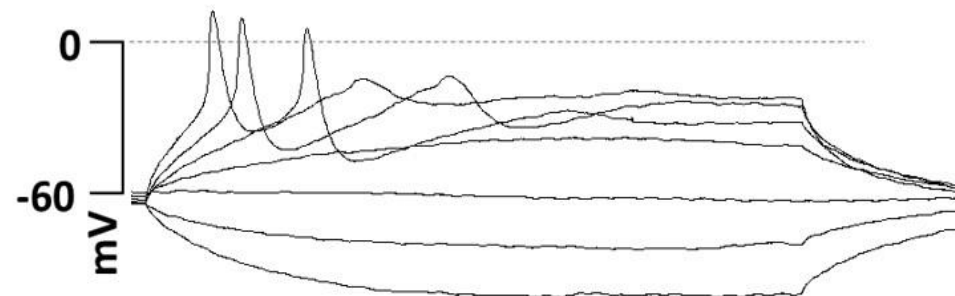


Evoked currents



SH-SY5Y differentiated cells

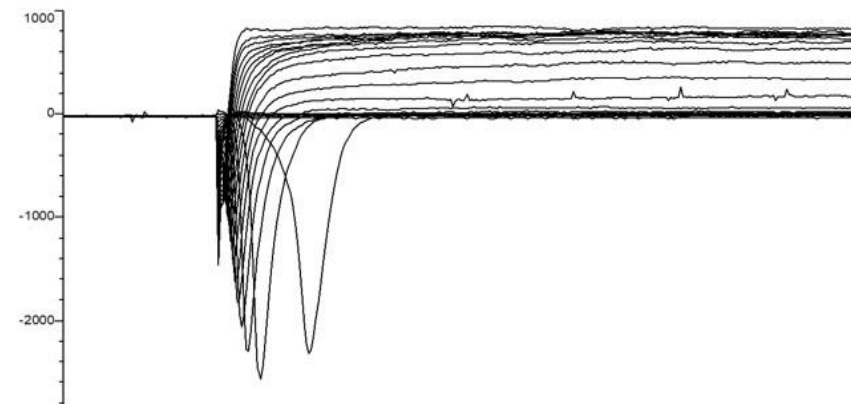
Evoked Action Potential



Resting Potential



Evoked currents



GRAZIE PER LA VOSTRA ATTENZIONE 😊