SUPPRESSION OF PHYTOPATHOGENIC FUNGI BY PSEUDOMONAS SSP. STRAINS

Wojciech Sokołowski, Magdalena Karaś, Sylwia Wdowiak-Wróbel, Monika Marek-Kozaczuk, Michał Kalita, Karolina Włodarczyk Department of Genetics and Microbiology, Maria Curie-Sklodowska University in Lublin, Poland

Introduction

Among many modern and eco-friendly approaches in the plant health management, use of biopesticides, containing living microorganisms or their metabolites, seems a more sustainable and environmentally friendly way of plant disease control in crop production [Ram et al., 2018]. Among the various biocontrol agents, the Pseudomonas species are the promising candidates for agricultural application, due to the wide spectrum of enzyme activities and antimicrobial molecules production [Sheoran et al., 2015].

Materials and methods

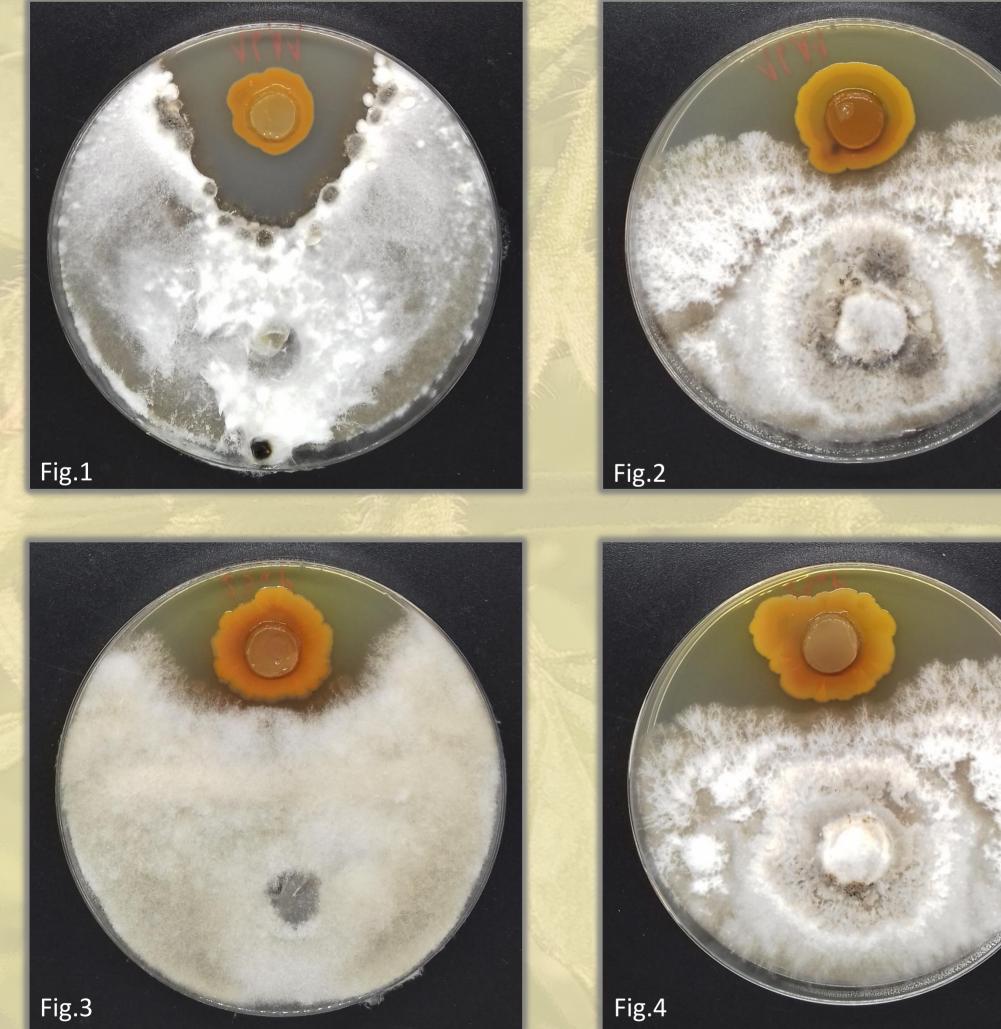
Four Pseudomonas strains (16A1, 16B1, 23aP and 154), isolated from root nodules of wild leguminous plants growing in the southeastern part of Poland were characterized. Antagonism of bacterial isolates toward phytopathogenic fungi Botrytis cinerea, Diaporthe rudis, Fusarium equiseti, Fusarium oxysporum and Sclerotinia sclerotiorum was performed in vitro by the disc diffusion method using the potato dextrose agar (PDA) plates [Balouiri et al., 2016]. We also assessed their plant-growth promoting properties including production of siderophores, salicylic acid and hydrogen cyanide (HCN) [Arnow, 1937; Lorck, 1948].

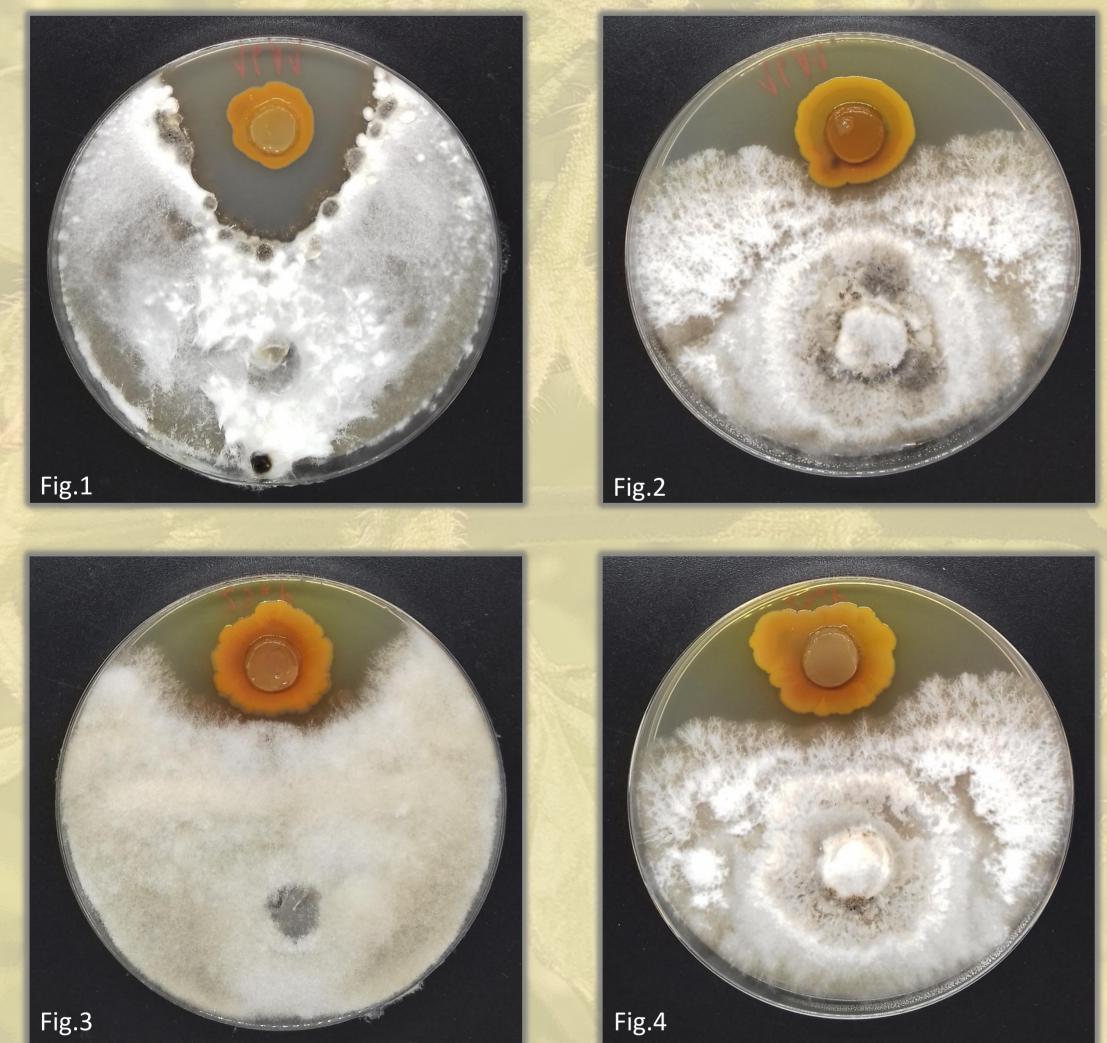
Suppression range of phytopathogenic fungi

Diaporthe Sclerotinia Botrytis Fusarium Fusarium rudis cinerea equiseti sclerotiorum oxysporum

Results

All the studied strains inhibited the growth of the phytopathogenic fungi on PDA agar plate assay (Fig. 1-4), but the degree of inhibition varied per endophyte. Strains 23aP and 16A1 were the most effective against phytopathogenic fungi, especially against the white mold disease agent, Sclerotinia sclerotiorum. Moreover, strain 16A1 inhibited the growth of all phytopathogenic fungi used in this assay (Fig. 5).





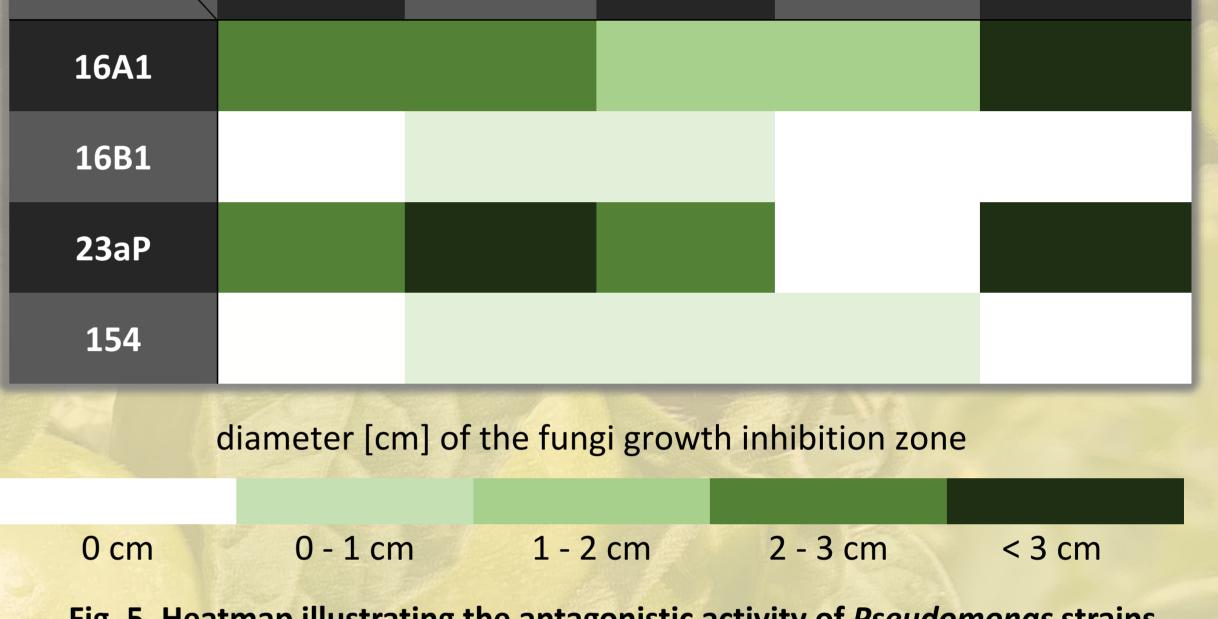


Fig. 5. Heatmap illustrating the antagonistic activity of *Pseudomonas* strains against phytopathogenic fungi

The results obtained showed that all tested strains were able to produce molecules with biocontrol potential as: hydroxamate siderophores, salicylic acid and hydrogen cyanide however, to a different degree. The strain 16A1 showed the highest level of hydroxamate siderophores production and a relatively high level of salicylic acid synthesis. While, strain 154 indicated the strongest reaction in the hydrogen cyanide test (Fig. 6).

Salicylic acid

Siderophore [ug/ml]

Hydrogen Cyanide

Fig. 1-4. The inhibition of phytopathogenic fungi growth after 14 days of incubation on PDA medium

Fig. 1. strain 16A1 and the growth inhibition zone of *S. sclerotiorum*; Fig. 2. strain 16A1 and the growth inhibition zone of *D. rudis*; Fig. 3. strain 23aP and the growth inhibition zone of *B. cinerea*; Fig. 4. strain 23aP and the growth inhibition zone of *D. rudis*

	[µg/ml]	[µg/ml]	+ weak, ++ medium, +++ strong reaction
16A1	0,75	23	+
16B1	0,55	31	+
23aP	0,19	22	++
154	0,08	6	+++

Fig. 6. The production of hydroxamate siderophores, salicylic acid and hydrogen cyanide by tested *Pseudomonas* strains

Conclusions

Taking into account results of *in vitro* antagonistic assays observed as growth suppression of some phytopathogenic fungi, it can be concluded, that all studied strains has biocontrol potential and they seem to be a promising candidates to use in sustainable agriculture. However, due to no clear correlation was found between the production level of the tested metabolites and antifungal activity, it is not possible to indicate mechanisms underling these phenomena. It is likely that the synergistic action of these and other untested mechanisms contributed to observed results, therefore further investigations needs to be done.

1. Arnow L., 1937. Colorimetric determination of the components of 3,4-dihydroxyphenylalaninetyrosine mixtures. J. Biol. Chem. 118, 531-537. doi:10.1016/s0021-9258(18)74509-2;

2. Balouiri M., Sadiki M., Ibnsouda S.K., 2016. Methods for in vitro evaluating antimicrobial activity: A review. J. Pharm. Anal. 6(2), 71-79. doi:10.1016/j.jpha.2015.11.005;

3. Lorck H., 1948. Production of hydrocyanic acid by bacteria. Physiol. Plantarum 1, 142-146;

4. Ram R.M., Keswani C., Bisen K., Tripathi R., Singh S.P., Singh H.B., 2018. Biocontrol Technology. Omics Technologies and Bio-Engineering, 177–190. doi:10.1016/b978-0-12-815870-8.00010-3; 5. Sheorana N., Nadakkakathb A.V., Munjala V., Kunduc A., Subaharand K., Venugopald V., Rajammab S., Eapenb S.J., Kumar A., 2015. Genetic analysis of plant endophytic Pseudomonas putida BP25 and chemo-profiling of its antimicrobial volatile organic

compounds. Microbiol. Res. 173, 66–78. doi: 10.1016/j.micres.2015.02.001.